

The influence of the sample preparation of carrots (*Daucus Carota* L. *Neptun*) on the antioxidant activity and phenolic compounds

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The growing demand for organic food in the world requires the assessment of the value aspects of food quality, its safety, nutritional content and biological-physiological process.

Fruits and vegetables are good sources of natural antioxidants containing many different antioxidant components. These antioxidants include carotenoids, vitamins, phenolic compounds, flavonoids, dietary glutathione and endogenous metabolites which have been shown to function as singlet and triplet oxygen quenchers, free radical scavengers, peroxide decomposers, enzyme inhibitors, and synergists. Phenolic compounds are found in most fruits and vegetables (Zhang, Hamauzu, 2005). Carrots and their fresh produce (shredded carrots, sliced carrots and carrot juice) may protect humans against certain types of cancer and cardiovascular diseases (Krinsky, Johnson, 2005). Carrots contain mainly hydroxycinnamic acids and derivatives. Among them, chlorogenic acid is a major hydroxycinnamic acid, representing from 42.2% to 61.8% of total phenolic compounds (Wang et al., 2006). In addition to the above mentioned compounds found in natural foods, vitamins C and E, beta-carotene and tocopherol are known to possess antioxidant potential.

The aim of the research was to determine the total antioxidant activity and phenolic compounds which operate as free radical scavengers, peroxide decomposers, enzyme inhibitors and synergists. Carrot breed *Neptun* was obtained from Lithuanian Institute of Horticulture. Correlations between antioxidant activity and phenolic compounds were determined by using three drying methods (carrots were sliced and dried in desiccator at +40 °C and lyophilised under vacuum conditions (freeze-dried) by –72 °C for sample preparation. Antioxidant activity was determined by using DPPH radicals spectrophotometrically. The influence of drying method was not significant $p > 0.05$, but solvent concentration showed significant correlation with antioxidant activity and phenolic compound ($p < 0.05$). Among the weight of raw material and various ratios of methanolic extracts, significantly the best antioxidant activity was shown by 0.5 g and 75% of sliced desiccated, freeze-dried and pressed desiccated samples. The total phenolic content of carrot material

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investigated during this research varied from 0.07 to 0.31 mg/ml of the methanolic carrot extracts and from 1.51 to 7.01 mg / g of dry carrot content, expressed by gallic acid equivalent (GAE). The time effect for DPPH scavenging was significant until 30 min ($p < 0.05$). If the reaction achieved a plato effect (balance), approx. 80% of antioxidant activity, the time effect was not significant ($p > 0.05$).

Key words: *Daucus carota* L. *Neptun*, phenolics, antioxidant activity, methanolic extracts

INTRODUCTION

The presence of phenolic compounds in carrots contributes to their sensory qualities, like colour (Zhang et al., 2004), bitterness (Kreutzmann et al., 2008), or aroma (Naczka, Shahidi, 2003). Therefore, the response of phenolic compounds could be used as a good indicator to evaluate the vegetables quality during processing and storage. The quality of vegetables mostly depends on the components accumulated in fresh matrix, peel and cortex (Goncalves et al., 2010). Therefore it is very important to precisely estimate the quality of carrots. There were determined relative levels of antioxidant activity in several of carrots consisting of a variety of vegetable families, selected on the basis of their widespread use in traditional consumption via scavenging of 2,2-diphenyl-2-picrylhydrazyl hydrate (DPPH) radical determined spectrophotometrically. A number of carrot species with appreciable levels of antioxidant activity against the DPPH radical were identified as potential sources of free radical scavenging compounds (Puodžiūnienė et al., 2005).

Active compounds

Fruits and vegetables are good sources of natural antioxidants containing many different antioxidant components. These antioxidants include carotenoids, vitamins, phenolic compounds, flavonoids, dietary glutathione and endogenous metabolites and have been shown to function as singlet and triplet oxygen quenchers, free radical scavengers, peroxide decomposers, enzyme inhibitors, and synergists (Larson, 1988). Phenolic compounds are found in most fruits and vegetables. Carrots contain mainly hydroxycinnamic acids and derivatives. Among them, chlorogenic acid is a major hydroxycinnamic acid, representing from 42.2% to 61.8% of total phenolic compounds (Wang et al., 2006). In addition to the above mentioned compounds found in natural foods, vitamins C and E, beta-carotene and tocopherol are known to possess antioxidant potential (Prior, 2003).

Carrot breed (*Daucus carota* L. *Neptun*) was chosen for the experiment from Lithuanian Institute of Horticulture, Babtai, Lithuania. Correlations between some of radical scavenging parameters were determined.

The aim of the research was to evaluate the drying method and sample preparation to determine antioxidant activity (DPPH) and total phenolic compounds. The task was to evaluate sample preparation of carrots by using three drying methods and different ratio (75% and 100%) of methanol for carrot samples extractions (freeze-dried under vacuum condition at $-72\text{ }^{\circ}\text{C}$, the second – sliced with blender and the third – pressed carrots by the juice press, then desiccated in the fruit and vegetable desiccator at $+40\text{ }^{\circ}\text{C}$).

MATERIALS AND METHODS

Materials

Fresh carrots (*Daucus carota* L. *Neptun*) were provided by Lithuanian Institute of Horticulture, Babtai, Lithuania. Methanol (CH_3OH) was of analytical grade (Sigma-Aldrich, Germany), 2,2-diphenyl-2-picrylhydrazyl hydrate (DPPH) was from Sigma-Aldrich (Germany). Bidistilled water was used for the experiments, (2N) Folin-Ciocalteu reagent was obtained from Sigma (USA), Na_2CO_3 p.a. (Chempur, Poland).

Preparation of carrot extracts in methanol

Sample preparation is the first crucial step in the analysis. In this research, sample preparation is similar to the preparation of medicinal herbs. Fresh carrots were sliced by “BRAUN” blender (Germany) and dried at $+40\text{ }^{\circ}\text{C}$ in desiccator (Scarlett SC-420), half of them were freeze-dried under vacuum conditions at $-72\text{ }^{\circ}\text{C}$ up to 72 h (Scanvac coolSafe™ 55-9, UK) and the third drying method involved carrots pressed by juice press Phillipps (Germany) and dried at $+40\text{ }^{\circ}\text{C}$ in desiccator (Scarlett SC-420). Twelve liquid maceration procedures were applied by shaking flasks with 0.5 g and 0.25 g of finely grounded dried carrots and different percent of solvent (75% and 100% CH_3OH) in a GFL3017 shaking machine. Extraction step lasted 1 h.

Evaluation of antioxidant activity by means of DPPH radical scavenging assay

Among chemical methods applied to determine the antioxidant activity of a compound, DPPH• (2,2-diphenyl-1-picrylhydrazyl) is one of the most frequently used methods because it is practical, fast and stable (Espin et al., 2000). Radical scavenging activity of methanolic carrot extracts against stable DPPH was determined spectrophotometrically. When DPPH reacts with an antioxidant compound, which can donate hydrogen, it is reduced. The changes in colour (from deep violet to light yellow) were measured at 515 nm on UV/visible light spectrophotometer T70+UV/VIS Spectrometer PG Instruments Ltd. Radical scavenging activity of extracts was measured by a slightly modified method of Brand-Williams et al., 1995 as described below. The solution of DPPH in methanol was prepared daily, prior to UV measurements. 3 ml of this solution were mixed with 77 μl of extract solution in 1 cm path length cuvettes. The samples were kept in the dark for 15 min at ambient temperature and then the decrease in absorption was measured. Absorption of blank sample containing the adequate amount of methanol and DPPH solution was prepared and measured daily. The experiment was carried out in triplicate. Radical scavenging activity was calculated by the following formula:

$$\% \text{Inhibition} = \frac{\text{AB} - \text{AA}}{\text{AB}} * 100\%$$

where: AB – absorption of blank sample ($t = 0$ min); AA – absorption of tested extract solution ($t = 15$ min, $t = 30$ min, $t = 45$ min). All assessments were performed in triplicate.

The content of phenolic compounds was determined by Folin-Ciocalteu colorimetric method (Folin-Ciocalteu, 1927; Wu Cu et al., 2007). The total phenolic content in carrot extracts was analyzed spectrophotometrically on UV/visible light spectrophotometer T70+UV/VIS Spectrometer PG Instruments Ltd using a modification of Folin-Ciocalteu

colorimetric method (Folin-Ciocalteu 1927; Singleton, Rossi, 1965). This method consists of determining the total polyphenol content through oxidation of phenolic compounds using a mix of phosphotungstic and phosphomolybdic acids in base medium, producing blue acids of tungsten and molybdenum. Absorbance of these acids was then read at 765 nm (Folin, Ciocalteu, 1927). The results were expressed in mg gallic acid / 100 mL. Each assay was performed in triplicate. 0.05 mL of methanolic carrot extracts were mixed with 1.25 mL of double-distilled water, followed by the addition of 0.05 mL (2N) Folin-Ciocalteu reagent, and then well mixed. After being allowed to stand for 6 min 0.25 mL of 20% sodium carbonate solution was added. The colour was developed by keeping at room temperature for 30 min, and the absorbance measured at 760 nm using UV visible spectrophotometer. The measurement was compared to a standard curve prepared using gallic acid solution and expressed as the mean, in mg, calculated from equation $y = 0.0931x + 0.0134$ ($R^2 = 0.9616$) of gallic acid equivalent (GAE) per g of carrot dry material from triplicate extracts (Prior, 2003).

Statistical analysis

Data was analysed by SPSS statistic program (Portable_SPSS_v19.0.0.329), the disperse analysis method, Spearman correlation was obtained between antioxidant activity and phenolic compounds, time effect for antioxidant activity, dry mass of carrots content, influence of solvent percentage and, finally, the drying methods for carrots. Statistical analysis was performed using correlation matrixes test and $p < 0.05$ was used as the level of significance: one-way analysis of variance (ANOVA) followed by Tuckey's post test were used to assess significant differences ($p < 0.05$) in the same groups. The test of homogeneity was $p < 0.05$ value. The assays of total antioxidant activity, amount of phenolic compounds were carried out in triplicate. The values (% and mg/g) were presented as means \pm standard deviation (SD) in Figs. 1–2, correspondingly.

RESULTS AND DISCUSSION

Determination of antioxidant activity and phenolic compounds by various sample preparations of carrot breed *Neptun* is described

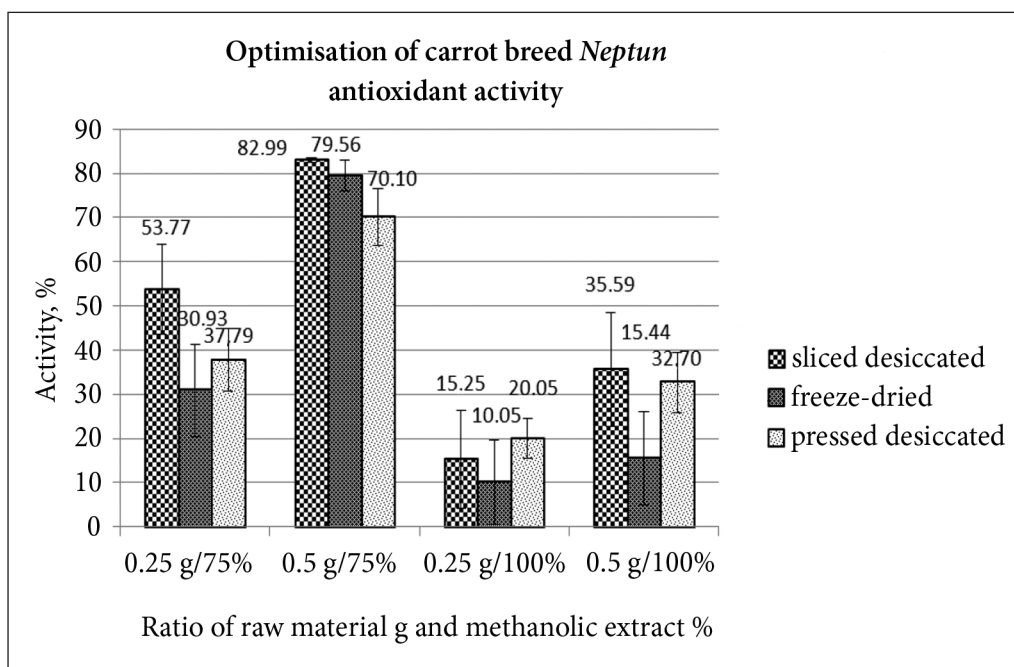


Fig. 1. DPPH radical scavenger activity, %

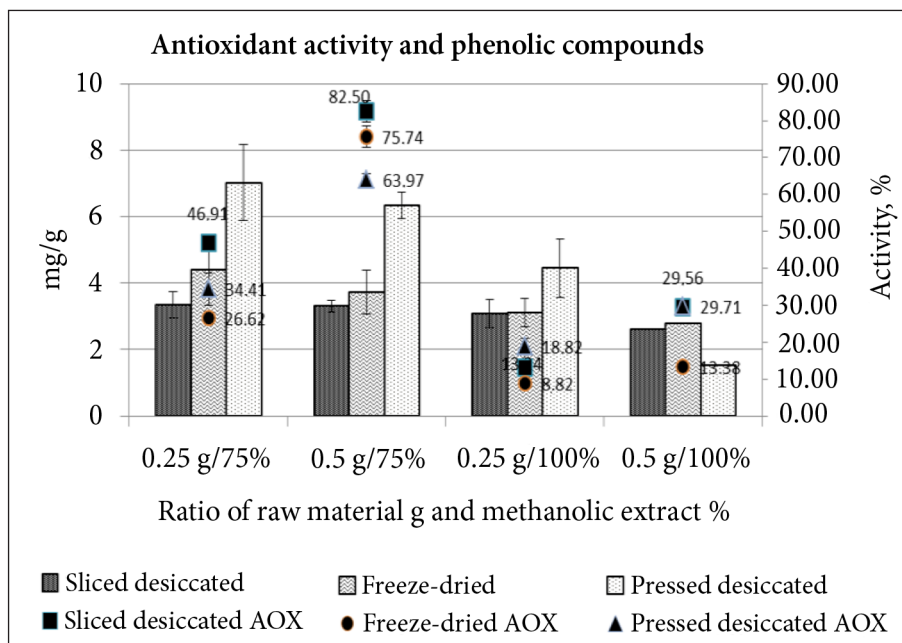


Fig. 2. Content of phenolic compounds, using gallic acid as a standard and expressed as mg/g of gallic acid equivalent (GAE) in dry carrot material (Folin-Ciocalteu method). Antioxidant activity % is shown as a point of AOX

below. The sample preparation is a crucial step of biocompounds determination in the plant analysis. Sample preparation for carrots is similar to that as for medicinal herbs. Methanolic extract was obtained using 0.5 g and 0.25 g of dry carrot material and 10 ml 75% and 100% of methanol. Results are shown in Fig. 1.

Antioxidant activity was determined using DPPH radicals. Influence of drying method (sliced and dried in desiccator at +40 °C or lyophilised under vacuum conditions (freeze-dried) at -72 °C) was not significant $p > 0.05$, but solvent concentration showed significant correlation with antioxidant activity and phenolic compound ($p < 0.05$). Among the weight of raw material and various ratios of methanolic extracts, significantly the best antioxidant activity was shown by 0.5 g and 75% of sliced desiccated, freeze-dried and pressed desiccated samples (Fig. 1). According to literature, phenolic compounds are mostly responsible and correlate with antioxidant activity. Polarity of solvent has great influence on the extracted compounds and it depends on the dielectric constant of solvent mixture.

“Solvent(s)” means a substance or substances in which various other substances may be fully or partially dissolved. Preferred solvents include aqueous solvents, and solvents having a dielectric constant less than that of water. The solvents water, methanol, ethanol, 1-propanol, 1-butanol, and acetic acid are polar protic solvents having a hydrogen atom attached to an electronegative atom, typically oxygen. Solvents having a dielectric constant less than that of water are particularly useful in the formation of the inventive complexes. In literature, dielectric constant of water is 80 and that of methanol – 33 (Guittard et al., 2005).

The total phenolic content of carrot material investigated varied from 0.07 to 0.31 mg / ml of the methanolic carrot extracts and from 1.51 to 7.01 mg/g of dry carrot content expressed by gallic acid equivalent (GAE). Antioxidant activity % is shown as a point of AOX. Extracts of 0.25 g of pressed carrots showed up to 7.01 mg/g phenolic content in dry carrot material with 75% of methanol solvent. Ratio of solvent was significant for pressed carrots ($p < 0.05$). The amount of phenolic compounds showed mean correlation ($R^2 = 0.608$) to the

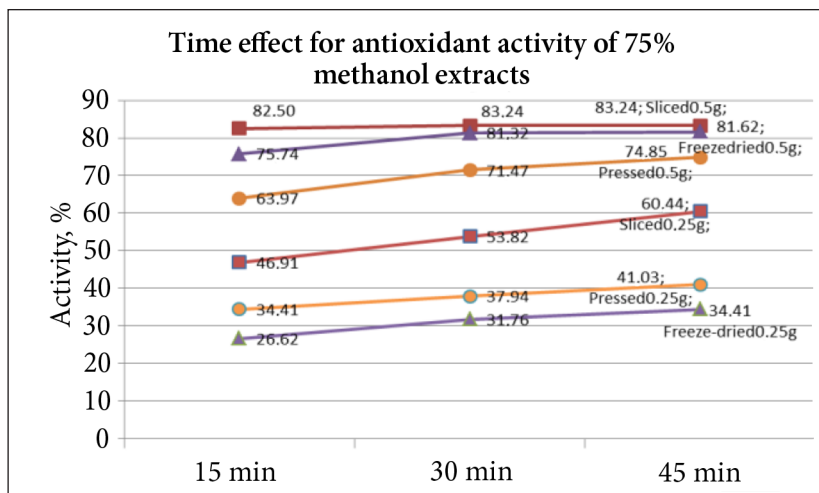


Fig. 3. The time impact on the dynamic of the antioxidant activity (by 75% methanolic extracts)

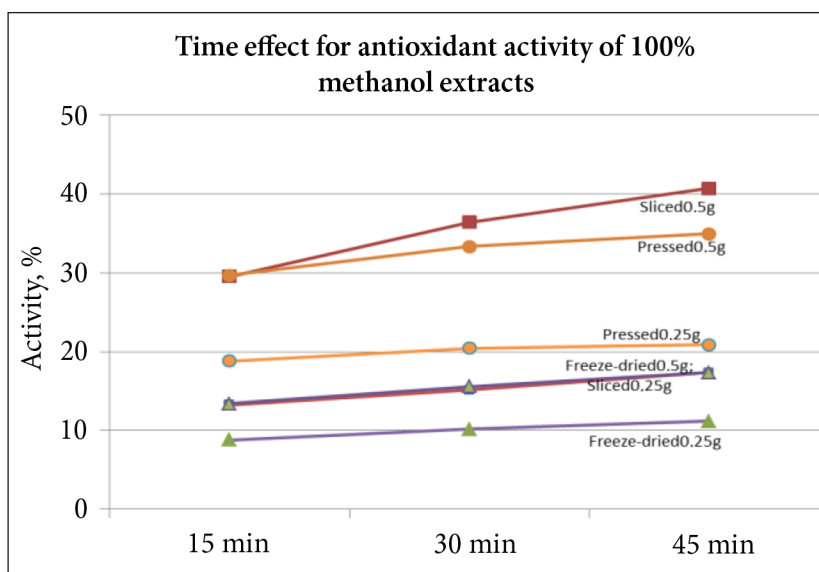


Fig. 4. Dynamic of AOX activity in 100% extracts

antioxidant activity. Correlation is significant at 0.05 level (2-tailed).

The dynamic reaction of antioxidant activity for the time period was evaluated and absorbance was read at 515 nm at different time intervals until the reaction reached stability. According to literature, most often the value of absorbance is recorded after 15 min. The results are shown in Fig. 3.

Significant time effect was until 30 min, $p < 0.05$. If the reaction achieved a plato effect (balance), about 80% of antioxidant activity (sliced carrots 0.5 g), the time effect was not

significant ($p > 0.05$). The best result until the reaction reached stability was achieved using 0.5 g of carrot material with 75% of solvent.

Carrot extracts made with 100% methanol have significant balance, if 0.25 g material was taken. In most cases the time effect was significant by sliced 0.5 g and pressed 0.5 g carrots, $p < 0.05$, until 45 min, but no significant change was for the remaining $p > 0.6$. The reaction achieved a plato effect (balance) approx. 10–20% of antioxidant activity, the time effect was not significant ($p > 0.05$) by pressed 0.25 g, freeze-dried 0.25 g and 0.5 g, and sliced

0.25 g. In this case, no changes in reaction dynamics were noticed by low antioxidant activity, but over 10% changes of antioxidant activity values were noticed in case of the sliced 0.5 g and pressed 0.5 g carrots. According to literature, it depends on dissolved substances in solvent from carrots.

CONCLUSIONS

Antioxidant activity was determined using DPPH radicals. Influence of drying method (sliced and dried in desiccator at +40 °C or lyophilised under vacuum conditions (freeze-dried) at -72 °C) was not significant $p > 0.05$, but solvent concentration showed significant correlation with antioxidant activity and phenolic compound ($p < 0.05$). Among the weight of raw dried material and the various ratios of methanolic extracts, significantly the best antioxidant activity was shown by 0.5 g and 75% of sliced desiccated, freeze-dried and pressed desiccated samples. For the next step of sample preparation, extraction with 75% of solvent and 0.5 g of dry material of carrots would be preferable. Drying methods had no significant value for antioxidant activity ($p > 0.05$), but were significant for phenolic compounds ($p < 0.05$).

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- tumai nereikšmingi laiko atžvilgiu. 0,5 g kapotose morkose, džiovintose vaisių džiovyklėje, pasiekus 80 % aktyvumą, reakcija pasiekdavo pusiausvyrą. Džiovinimo poveikis antioksidaciniam aktyvumui buvo nedidelis ($p > 0,05$), bet reikšmingas bendram fenolinių junginių kiekiui ($p < 0,05$).

Raktažodžiai: *Daucus carota* L. 'Neptun', fenoliniai junginiai, antioksidacinis aktyvumas, metanolio ekstraktas

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**ĮPRASTINIŲ MORKŲ (*DAUCUS CAROTA* L.)
BANDINIŲ DŽIOVINIMO POVEIKIS
ANTIOKSIDACINIAM AKTYVUMUI IR
FENOLINIŲ JUNGINIŲ KIEKIUI**

Santrauka

Skirtingomis ekstrakcijos sąlygomis – derinant sausos masės kiekį ir tirpiklio procentinį santykį, atliekant reakcijas su laisvuju radikalu DPPH bei matuojant spektrofotometriškai ties 515 nm bangos ilgiu – siekta nustatyti antioksidacinį 'Neptun' morkų veislės aktyvumą. Šioje morkų veislėje rasti fenolinių junginių kiekiai svyravo tarp 0,07 ir 0,31 mg/ml metanolio ekstrakto ir tarp 1,51 ir 7,1 mg/g sausos morkų masės pagal galo rūgšties ekvivalentus (Folin-Ciocalteu modifikuotas metodas). Didžiausias antioksidacinis aktyvumas – 82 % nustatytas sukapotų ir vaisių džiovyklėje išdžiovintų morkų, kai 0,5 g sausos morkų žaliavos ekstrahuota 75 % metanolio. Mažiausias aktyvumas – apie 8 % gautas ekstrahuojant 0,25 g sausos morkų žaliavos 100 % metanolio tirpikliu. Išmatavus antioksidacinio aktyvumo kaitą laiko atžvilgiu kas 15min., reikšmingi skirtumai gauti iki 45min. taikant 100 % metanolio ekstrakciją 0,5 g kapotai sausai medžiagai ($p > 0,05$). Su 75 % metanolio ekstrakcija gauti skir-